In the Specification

Please replace the paragraph found on page 20, line 2 through page 21, line 2 of the specification with the following paragraph:

BCL-2 amplification is performed essentially as described by Tu Y et al., *Upregulated expression of Bcl-2 in multiple myeloma cells induced by exposure to doxorubicin, etoposide, and hydrogen peroxide*, Blood 88:1805,1996. Briefly, 20 μL of PCR reaction mixture (IX PCR buffer, 50 pmol of BCL-2 specific amplimers, 0.25 U Taq polymerase [Boehrin.-erMannheim], 1.25 μCi [α-^{32}p]-dCTP) is added to 5 μL cDNA, followed by incubation at 94°C for 5 minutes and then 26 cycles of 94°C for one minute, 72°C for one minute, and a final extension at 72°C for five minutes in a thermal cycler (Perkin-Elmer Cetus). Histone 3.3 is amplified as described by Futscher Bwet al., *Quantitative polymerase chain reaction analysis of MDR1 mRNA in multiple myetoma cell lines and clinical specimens*, Anal Biochem 213:414, 1993, and is used as a control for RNA integrity and quantity. Bcl-XI and -Xs are amplified essentially as described by Benito A, et al., *Apoptosis induced by erytheroid differentiation of human leukemia cell lines is inhibited by Bct-XL*, Blood 87:3837, 1996, using 26 cycles of PCR. The 258 base pair BAX amplicon is amplified using the following primers (Biosynthesis, Lewisville, Tex.) and conditions: BAX-upstream (5'-ACCAAGAAGCTGAGCGAGTGTC- TC-3') (SEQ ID NO:1), BAXdownstream (5'-CAATGTCCAGCCCATGATGG-3') (SEQ ID NO:2), cDNA denaturation for one minute at 94°C, annealing for 15 seconds at 60°C, primer extension for 15 seconds at 72°C, with a final extension for 5 minutes. All samples are loaded on a five percent nondenaturing polyacrylamide gel and electrophoresed for two hours at 80V. For determination of incorporated radionucleotide, gels are dried down and exposed to a phosphoroimaging plate (Molecular Dynamics, Inc) overnight. Plates are then scanned on a phosphorimager (Molecular Dynamics) and band intensities (pixels/unit area) for Bcl-2, Bcl-XI, Bcl-Xs, and BAX are analyzed normalized to Histone 3.3 expression. PCR amplification of the MDRI, MRP, and LRP genes is performed essentially as described by Abbaszadegan M. et al., *Analysis of multidrug resistance-associated protein (MRP) messenger RNA in normal and malignant hematapoietic cells*, Cancer Res 54:4676,1994, and Komarov P. et al., *Activation of the LRP Oun-resistance-associated protein) gene by short-term exposure of human*

leukemia cells to phorbol ester and cytarabine, Oncology Res 10: 1 85, 1998, by using the housekeeping genes histone 3.3 (MDRI) or β-actin (MRP and LRP) as internal standards. cDNA synthesized from 8226/DOX6 RNA is used as a positive control for MDRI PCR. For all reactions, optimal cycle numbers are used and are within the exponential range of PCR amplification as determined by previous experiments.

Please replace the paragraph found on page 33, lines 8-25 of the specification with the following paragraph:

Total cellular RNA is collected from $3x10^6$ cells using TRIzol reagent (Gibco). RNA is quantitated on a spectophotometer at 260 nm. cDNA is synthesized in a 20 ul reverse transcription reaction containing 100 ng RNA, 1 X PCR Buffer (10 mM Tris, pH 8.3-50 mMKCL-1.5 mM $MgCl_2$), 1 mM concentrations each of dATP, dGTP, dCTP, and dTTP; 100 pmol random hexamers, 20 units RNAse inhibitor, and 6 units avian megalovirus reverse transcriptase (Boeringer-Mannheim). The reaction is prepared on ice then incubated at 42°C for 1 hour, 99°C for 10 minutes, and quick chilled to 4°. The following primers are synthesized (Gibco/BRL) and used for α4-specific PCR reactions: 5'-ATGGCTCCCAATGTTAGTGTGG-3' (upstream) (SEQ ID NO:3) and 5'-CACTGGCTTCTTTTCCACTTTCC-3' (downstream) (SEQ ID NO:4). The 292 base pair α4 product is amplified using 20 μl of PCR reaction mixture (1X PCR buffer, 6.25 pmol of α4 specific amplimers), 0.5 units Taq polymerase (Boehringer-Mannheim), and 5 μl of cDNA. Samples are subjected to incubation at 94°C for 1 minute and then 34 cycles of 94°C (15 sec), 72°C (15 sec), and a final extension at 72°C for 1 minute in a thermal cycler (Perkin-Elmer Cetus). Histone 3.3 is amplified and used as a control for RNA integrity and quantity. Samples are loaded on a 3 percent agarose gel, electrophoresed one hour at 80V, and visualized using EtBr staining. Restriction digests are used to confirm identity of products.

Please replace the paragraph found on page 42, lines 20-24 of the specification with the following paragraph:

A pre-established ~~microcrotiter~~ microtiter adhesion assay is used to determine if peptides such as RZ-3 are effective at preventing adhesion of myeloma cells to FN. The peptide RZ-3 contains all D-amino acids, and has the sequence:

krnviywkag (SEQ ID NO:6) (RZ-3)

in which the conventional one-letter code is used and lower case designates a D-amino acid, and the sequence is written from the N-terminus to the C-terminus.

Please add the following paragraphs at page 14, line 1 of the specification:

BRIEF DESCRIPTION OF THE SEQUENCES

SEQ ID NO:1 is an upstream primer.

SEQ ID NO:2 is a downstream primer.

SEQ ID NO:3 is an upstream primer.

SEQ ID NO:4 is a downstream primer.

SEQ ID NO:5 is the amino acid sequence Lys-Met-Val-Ile-Tyr-Trp-Lys-Ala-Gly.

SEQ ID NO:6 is the RZ-3 peptide, the amino acid sequence Lys-Met-Val-Ile-Tyr-Trp-Lys-Ala-Gly, wherein each amino acid is a D-amino acid.

Please insert the attached pages 1-3 of the Sequence Listing.